

# Report of the Third Biomedical Confidence Building Exercise

A. Alcaraz, A. Williams

April 30, 2013

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# Report of the Third Biomedical Confidence Building Exercise

Laboratory code: <u>07</u>

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# **SUMMARY: PARTICIPATING LABORATORY**

1. Participating laboratory

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laboratory/institute:	
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	USA
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Date of sample receipt:	February 5 <sup>th</sup> , 2013
Date of report:	April 30 <sup>th</sup> , 2013

#### 2. Analysts and authentication

	Name	Title	Signature
1	Armando Alcaraz	Pricipal Investigator	Company Change
2	Deon Anex	Research Scientist	Villing
3	Bradley Hart	FSC Director	Landhe.
4	Saphon Hok	Research Scientist	Loylus At
5	Carolyn Koester	Research Scientist	Carp Kan
6	Roald Leif	Research Scientist	Roal List
7	Brian Mayer	Research Scientist	BOXILE
8	Tuijuana Mitchell-Hall	QA Manager	Dujauna hitchell- Hall
9	Heather Mulcahy	Research Scientist	Steather Unically
10	Carlos Valdez	Research Scientist	Che fally
11	Alexander Vu	Research Scientist	aldi
12	Audrey Williams	Research Scientist	Andrew William

# **SUMMARY: QUALITY SYSTEM**

Quality system:
<ul> <li>☑ Described in a Quality Assurance Manual/Handbook. Quality system in accordance with:</li> <li>☑ ISO/IEC 17025</li> <li>☑ Other/none: (describe below)</li> </ul>
Accreditation accepted; Year of accreditation: 2001  Accreditation planned/pending; Year accreditation expected:
Accreditation body: <u>A2LA</u> Scope of accreditation: <u>CW analysis</u>
Summary for laboratories with either "other" accreditation or no planned accreditation:

# **SAMPLE SUMMARY: A**

Sample Code: P-301/07 Laboratory Assigned Code: CW-4-147-1

Description and condition of sample: Approximately 5 mL of plasma

No chemicals relevant to the purpose of analysis were found.

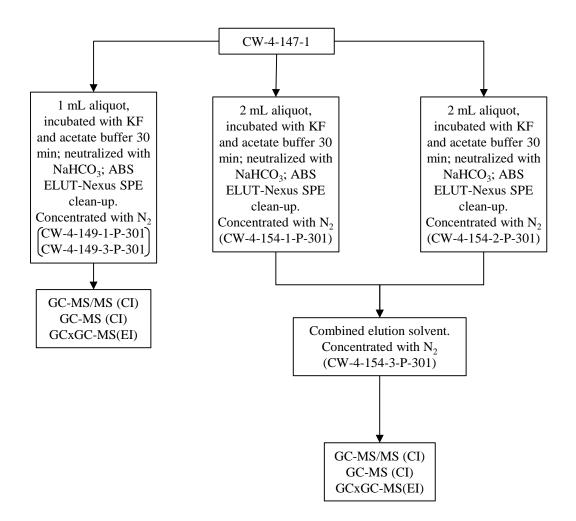
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# SAMPLE PREPARATION DESCRIPTION: A

1. Sample preparation

Initial Aliquot Code	Type of Sample Preparation	Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code
CW-4-147-1	Fluoride Reactivation	1 mL	Added 3 mL of acetate buffer and 220 μL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate and reduced sample volume to approximately 100 μL using nitrogen gas.	100 μL	CW-4-149-1-P-301
CW-4-149-1-P-301	Sample Split	50 μL	Split of sample CW-4-149-1-P-301.	50 μL	CW-4-149-3-P-301
CW-4-147-1	Fluoride Reactivation	2 mL	Added 3 mL of acetate buffer and 277 μL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate.	2 mL	CW-4-154-1-P-301
CW-4-147-1	Fluoride Reactivation	2 mL	Added 3mL of acetate buffer and 277 µL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate.	2 mL	CW-4-154-2-P-301
CW-4-154-1-P-301 CW-4-154-2-P-301	Concentrate combined samples	2 mLx2	Combined elution solvent from CW-4-154-1-P-301 and CW-4-154-2-P-301. Reduced sample volume to approximately 100 µL using nitrogen gas.	100 μL	CW-4-154-3-P-301

#### 2. Additional information



#### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by incubating the nerve agent, Sarin, with commercially procured human blood plasma. Cholinesterase activities of the plasma sample were measured using method outlined in Ellman (1961) before and after incubation to ensure agent-adduction occurred. The resultant agent-adducted plasma was used for subsequent method development.

Adducted plasma samples were reactivated using the process outlined in Degenhardt *et al* (2004). 1 mL of plasma was added to 3 mL of acetate buffer solution [0.189 M acetic acid; 10.8 mM sodium acetate]. Appropriate amount of Potassium Fluoride [4.51M] was added to achieve a final concentration of 0.25 M KF in the unknown sample work-up. Sample was allowed to incubate at room temperature for 30 minutes. 0.5 mL of 0.8 M sodium bicarbonate was added to neutralize acid.

Several methods were attempted to isolate the reactivated sarin from the sample:

- 1. Direct dichloromethane extraction
- 2. SPE preparation #1

Agilent ABS Elut-NEXUS (200 mg/6 mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

3. SPE preparation #2

Phenomenex Strata-X 33µm (30mg/3mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

An Agilent ABS Elut-Nexus SPE cartridge was conditioned, by gravity, with 2 mL of N-hexane, 2x2 mL ethyl acetate, followed by 2x2 mL of  $H_2O$ . 1 mL reactivated plasma sample was then loaded into cartridge and allowed to pass through. Sample was eluted with 2 mL of ethyl acetate into a clean vial. Sample was reduced in volume to approximately  $100 \, \mu L$  for analysis.

When the target compound was not detected in CW-4-147-1, a deviation was made to this method. CW-4-147-1 was reactivated by using two replicates of 2 mL instead of the 1 mL described above and the resulting eluted samples were combined for their respective samples to increase the analytical concentration of the target analyte, should it be present. The combined samples were then reduced in volume to approximately  $100~\mu L$  for analysis.

#### References

- G.L. Ellman. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharm 7: 88-95(1961).
- C. E. A. M. Degenhardt, K. Pleijsier, M. J. van der Schans, J. P. Langenberg, K. E. Preston, M. I. Solano, V. L. Maggio, J. R. Barr. Improvements of the Fluoride Reactivation Method for the verification of nerve agent exposure. Journal of Analytical Toxicology. 28(5): 364-371 (2004).

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# **SAMPLE SUMMARY: B**

Sample Code: P-302/07 Laboratory Assigned Code: CW-4-147-2

Description and condition of sample: Approximately 5 mL of plasma

No chemicals relevant to the purpose of analysis were found.

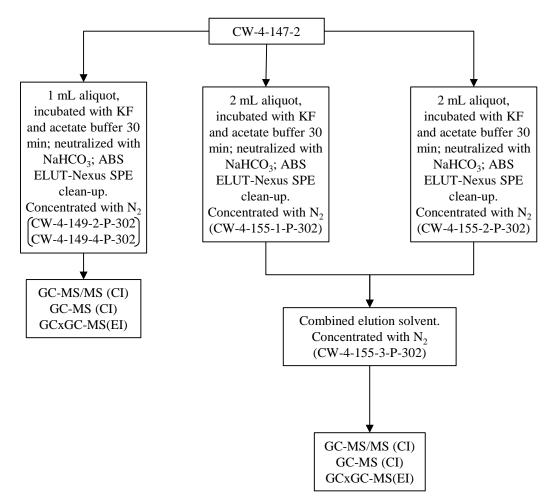
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# SAMPLE PREPARATION DESCRIPTION: B

1. Sample preparation

Initial Aliquot Code	<b>Type of Sample Preparation</b>	Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code	
CW-4-147-2	Fluoride Reactivation	1 mL	Added 3 mL of acetate buffer/0.25 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate and reduced sample volume approximately 100 µL with nitrogen gas.	100 μL	CW-4-149-2-P-302	
CW-4-149-2-P-302	Sample Split	100 μL	Split of sample CW-4-149-2-P-302.	50 μL	CW-4-149-4-P-302	
CW-4-147-2	Fluoride Reactivation	2 mL	Added 3 mL of acetate buffer and 277 µL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate.	2 mL	CW-4-155-1-P-302	
CW-4-147-2	Fluoride Reactivation	2 mL	Added 3 mL of acetate buffer and 277 µL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate.	2 mL	CW-4-155-2-P-302	
CW-4-155-1-P-302 CW-4-155-2-P-302	Concentrate combined samples	2 mLx2	Combined elution solvent from CW-4-155-1-P-302 and CW-4-155-2-P-302. Reduced sample volume to approximately 100 µL using nitrogen gas.	100 μL	CW-4-155-3-P-302	

#### 2. Additional information



#### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by incubating the nerve agent, Sarin, with commercially procured human blood plasma. Cholinesterase activities of the plasma sample were measured using method outlined in Ellman (1961) before and after incubation to ensure agent-adduction occurred. The resultant agent-adducted plasma was used for subsequent method development.

Adducted plasma samples were reactivated using the process outlined in Degenhardt *et al* (2004). 1 mL of plasma was added to 3 mL of acetate buffer solution [0.189 M acetic acid; 10.8 mM sodium acetate]. Appropriate amount of Potassium Fluoride [4.51M] was added to achieve a final concentration of 0.25 M KF in the unknown sample work-up. Sample was allowed to incubate at room temperature for 30 minutes. 0.5 mL of 0.8 M sodium bicarbonate was added to neutralize acid.

Several methods were attempted to isolate the reactivated sarin from the sample:

- 1. Direct dichloromethane extraction
- 2. SPE preparation #1

Agilent ABS Elut-NEXUS (200 mg/6 mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

3. SPE preparation #2

Phenomenex Strata-X 33µm (30mg/3mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

An Agilent ABS Elut-Nexus SPE cartridge was conditioned, by gravity, with 2 mL of N-hexane, 2x2 mL ethyl acetate, followed by 2x2 mL of  $H_2O$ . 1 mL reactivated plasma sample was then loaded into cartridge and allowed to pass through. Sample was eluted with 2 mL of ethyl acetate into a clean vial. Sample was reduced in volume to approximately 100  $\mu$ L for analysis.

When the target compound was not detected in CW-4-147-2, a deviation was made to this method. CW-4-147-2 was reactivated by using two replicates of 2 mL instead of the 1 mL described above and the resulting eluted samples were combined for their respective samples to increase the analytical concentration of the target analyte, should it be present. The combined samples were then reduced in volume to approximately  $100~\mu L$  for analysis.

#### References

- G.L. Ellman. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharm 7: 88-95(1961).
- C. E. A. M. Degenhardt, K. Pleijsier, M. J. van der Schans, J. P. Langenberg, K. E. Preston, M. I. Solano, V. L. Maggio, J. R. Barr. Improvements of the Fluoride Reactivation Method for the verification of nerve agent exposure. Journal of Analytical Toxicology. 28(5): 364-371 (2004).

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# SAMPLE SUMMARY: C

Sample Code: P-303/07 Laboratory Assigned Code: CW-4-147-3

**Description and condition of sample:** Approximately 5 mL of plasma

#### Chemical: C-1

Chemical name	& Structure	CAS#	Schedule
O-Isopropyl methylph	osphonofluoridate		
	107-44-8	1.A.01	
Aliquot(s)	Original/derivative	Analysis te	chnique
CW-4-150-4-P-303	Original	GC-MS/M	IS (CI)
CW-4-150-4-P-303	GC-MS (CI)		
CW-4-150-1-P-303	GCxGC-M	IS (EI)	
Comments:			

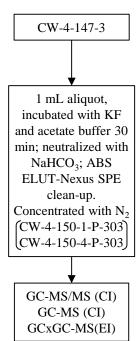
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# SAMPLE PREPARATION DESCRIPTION: C

1. Sample preparation

Initial Aliquot Code Type of Sample Preparation		Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code
CW-4-147-3	Fluoride Reactivation	1 mL	Added 3 mL of acetate buffer and 220 μL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate and reduced sample volume to approximately 100μL with nitrogen gas.	100 μL	CW-4-150-1-P-303
CW-4-150-1-P-303	Sample Split	50 μL	Split of sample CW-4-150-1-P-303.	50 μL	CW-4-150-4-P-303

•	A -	1.1:4:.		infa	rmation	
4.	A	ICHLLIC	шаі	11110	rmauon	



#### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by incubating the nerve agent, Sarin, with commercially procured human blood plasma. Cholinesterase activities of the plasma sample were measured using method outlined in Ellman (1961) before and after incubation to ensure agent-adduction occurred. The resultant agent-adducted plasma was used for subsequent method development.

Adducted plasma samples were reactivated using the process outlined in Degenhardt *et al* (2004). 1 mL of plasma was added to 3 mL of acetate buffer solution [0.189 M acetic acid; 10.8 mM sodium acetate]. Appropriate amount of Potassium Fluoride [4.51M] was added to achieve a final concentration of 0.25 M KF in the unknown sample work-up. Sample was allowed to incubate at room temperature for 30 minutes. 0.5 mL of 0.8 M sodium bicarbonate was added to neutralize acid.

Several methods were attempted to isolate the reactivated sarin from the sample:

- 1. Direct dichloromethane extraction
- 2. SPE preparation #1

Agilent ABS Elut-NEXUS (200 mg/6 mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

3. SPE preparation #2

Phenomenex Strata-X 33µm (30mg/3mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2mL of reactivated sample.

Elution: Ethyl Acetate

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

An Agilent ABS Elut-Nexus SPE cartridge was conditioned, by gravity, with 2 mL of N-hexane, 2x2 mL ethyl acetate, followed by 2x2 mL of  $H_2O$ . 1 mL reactivated plasma sample was then loaded into cartridge and allowed to pass through. Sample was eluted with 2 mL of ethyl acetate into a clean vial. Sample was reduced in volume to approximately  $100 \, \mu L$  for analysis.

#### **References**

- G.L. Ellman. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharm 7: 88-95(1961).
- C. E. A. M. Degenhardt, K. Pleijsier, M. J. van der Schans, J. P. Langenberg, K. E. Preston, M. I. Solano, V. L. Maggio, J. R. Barr. Improvements of the Fluoride Reactivation Method for the verification of nerve agent exposure. Journal of Analytical Toxicology. 28(5): 364-371 (2004).

## GC-MS/MS (CI) TECHNIQUE METHOD AND ANALYSIS DESCRIPTION

#### **Identification**

Chemical: C-1

**Aliquot code:** CW-4-150-4-P-303

**Datafile name:** TSQ0328

Compound identified as: Original Compound Compound reference: Reference Chemical (OPCW) Match algorithm and match factor: NIST, 980/999

#### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

**Column brand/phase:** Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 90 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

Solvent delay time: 3 min Electron energy: 100 eV Reaction gas: Ammonia Ionisation polarity: Positive

Scan range/time: 60-200 m/z in 1 second

Mass resolution: 0.7

Type of MS/MS scan: Product ion scan

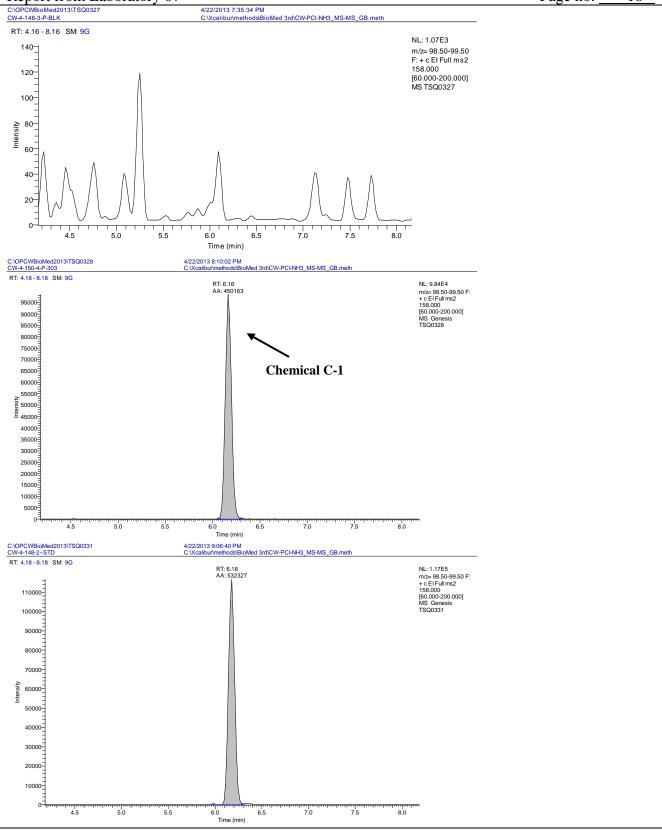
Precursor ion(s): m/z 158 Collision gas: Argon Collision Energy: 10

Tong/twomaitions	Sa	ample	Star	ndard	Criteria	D14
Ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
99	450163	1.0	532327	1.0	N/A	N/A
141	62037	0.138	57494	0.108	±30%	27.6%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

#### Remarks

<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\leq 10\%$ :  $\pm 50\%$ 

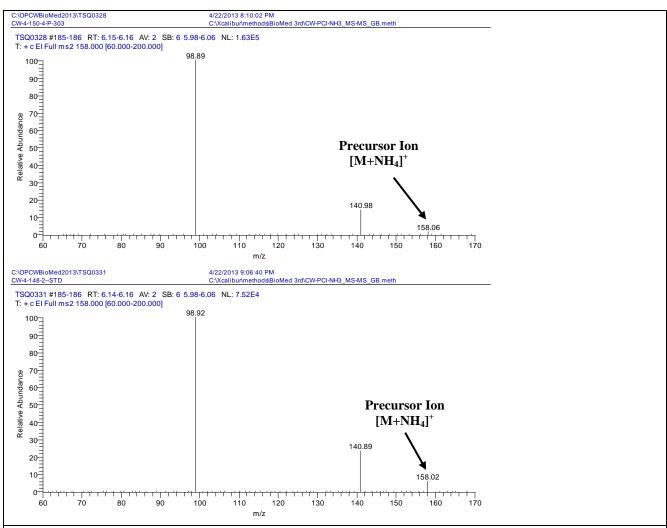


GC-MS/MS (CI) chromatograms supporting identification of Chemical C-1; EIC

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **O-Isopropyl methylphosphonofluoridate.** 



CI product mass spectra of:

Top: Chemical C-1 (aliquot code listed in header).

Bottom: Reference chemical of O-Isopropyl methylphosphonofluoridate.

## GC-MS(CI) TECHNIQUE METHOD AND ANALYSIS DESCRIPTION

#### **Identification**

Chemical: C-1

Aliquot code: CW000626.D

Datafile name: CW-4-150-4-P-303

Compound identified as: Original Compound

**Compound reference:** Reference Chemical (OPCW)

#### **Analysis Method**

GC Instrument manufacturer and type: Agilent 6890

Carrier gas: Helium

Flow control/rate: Constant Flow, 32 cm/sec Injection mode: Pulsed Splitless, 0.75 min

Injector temperature: 250 °C

**Column brand/phase:** Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 100 °C, 25 °C/min to 300 °C (3 min)

MS Instrument manufacturer and type: Agilent 5975 MSD

**Solvent delay time:** 5.4 min **Electron energy:** 82 eV

Scan range/time: SIM, m/z 141, 158, 100 µs dwell time

Source temperature: 230 °C

Reaction gas: Ammonia and flow: 21%

**Ionisation polarity:** Positive

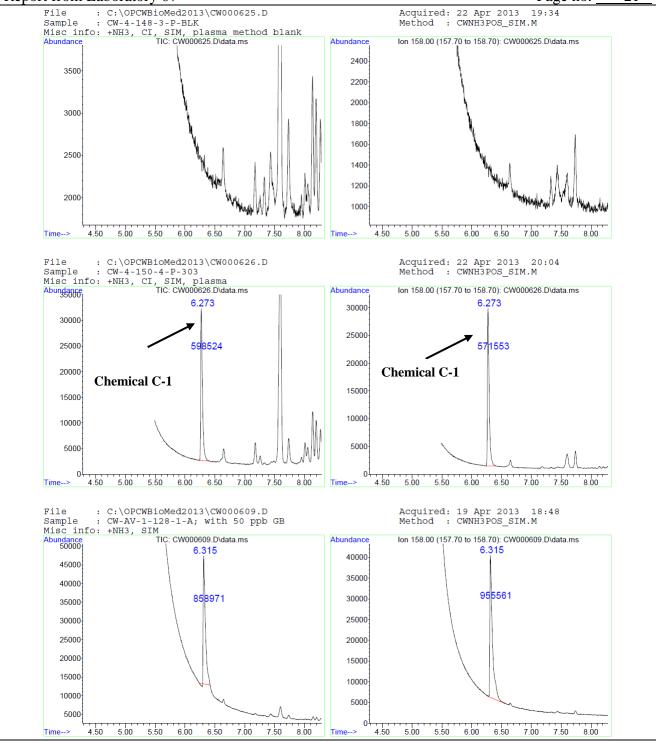
Ions/transitions	Sample		Standard		Criteria	Result
Tons/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
141	21504	0.038	28172	0.029	±50%	27.6%
158	571553	1.0	955561	1.0	N/A	N/A

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

#### **Remarks**

Due to the solvent delay of 5.4 min, the chromatogram could not be presented with a window of the retention time of the compound minus 2 min.

<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



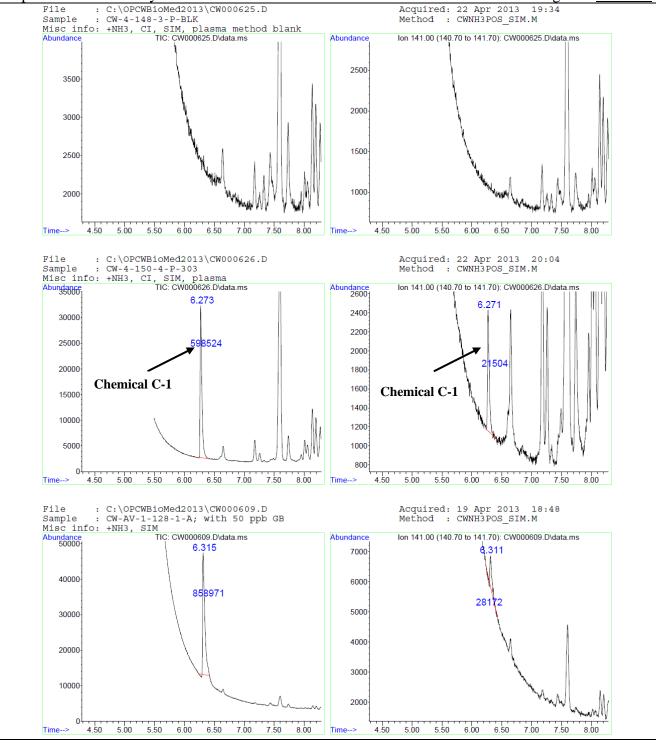
EI chromatograms supporting identification of Chemical C-1;

TIC on left, EIC (m/z 158) on right.

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **O-Isopropyl methylphosphonofluoridate**.



EI chromatograms supporting identification of Chemical C-1;

TIC on left, EIC (m/z 141) on right.

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of O-Isopropyl methylphosphonofluoridate.

### GCXGC-MS(EI) TECHNIQUE METHOD AND ANALYSIS DESCRIPTION

#### **Identification**

Chemical: C-1

**Aliquot code:** CW-4-150-1-P-303

**Datafile name:** TOF1335

Chemical identified as: Original Compound Chemical reference: Reference Chemical Match algorithm and match factor: 914/999

#### **Analysis Method**

GC Instrument manufacturer and type: Agilent 6890 GC

Carrier gas: Helium Flow rate: 1.10 mL/min

X Corrected constant flow via pressure ramps

**Injection mode:** Splitless, 0.17 min **Injector temperature:** 250° C

**Primary Column:** 

**Brand/phase:** Agilent HP-5 MS UI: (5% diphenyl 95% dimethyl polysiloxane)

**Column Length x ID x Film thickness:** 30 m x 0.25 mm x 0.25 µm **GC temperature programme:** 40°C (3 min), 8°C/min, 300°C (3 min)

**Secondary Column:** 

**Brand/phase:** Restek Rxi-17: (50%-Phenyl)-methylpolysiloxane **Column Length x ID x Film thickness:** 1.5 m x 0.18 mm x 0.18 μm **GC temperature programme:** 50°C (3 min), 8°C/min, 310°C (3 min)

X Modulator enabled

Modulator temperature offset: 20 °C, relative to the GC secondary oven temperature

**Modulation period:** 3.5 sec **Hot pulse time:** 0.6 sec

Cool time between stages: 1.15 sec

MS Instrument manufacturer and type: LECO Pegasus 4D

Solvent delay time: 3 minutes Detector voltage: 1750 V Electron energy: 70 eV Mass resolution: 0.6 u Scan range: 29-600 m/z Scan rate: 200 spectra/sec Source temperature: 250 °C

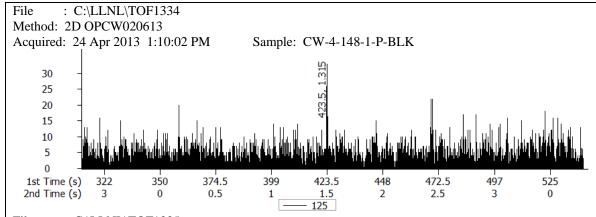
Ions/transitions	Sample		Standard		Criteria	Dogult	
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result	
39	3989.4	0.162	27388	0.152	±30%	6.6%	
81	3972.7	0.162	27547	0.153	±30%	5.5%	
99	24559	1.00	179664	1.00	N/A	N/A	
125	4239.4	0.173	36495	0.203	±25%	15.0%	

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

#### Remarks

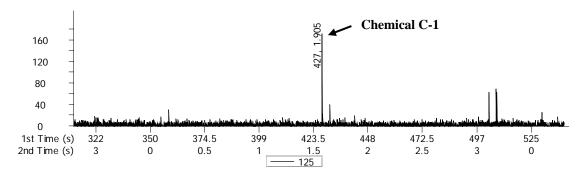
A small peak is present in the blank within 0.1 min of the target chemical. This peak has been tentatively identified as hexamethylcyclotrisiloxane through library matching (NIST, 960/999). The mass spectrum of this compound and the library spectrum are included.

<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\leq 10\%$ :  $\pm 50\%$ 



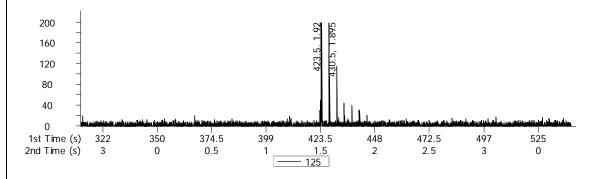
File : C:\LLNL\TOF1335 Method: 2D OPCW020613

Acquired: 24 Apr 2013 1:52:49 PM Sample: CW-4-150-1-P-303



File : C:\LLNL\TOF1336 Method: 2D OPCW020613

Acquired: 24 Apr 2013 4:34:02 PM Sample: CW-4-150-3-SARINSTD

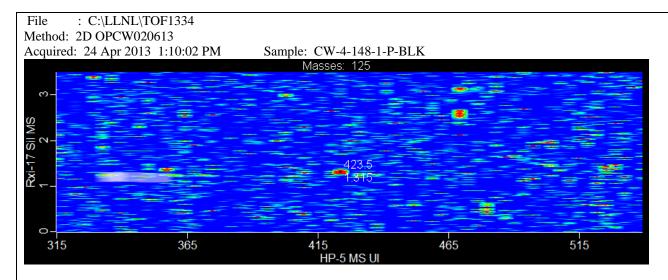


EI chromatograms supporting identification of **Chemical C-1**; EIC

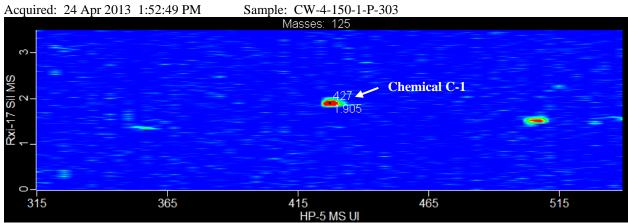
Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

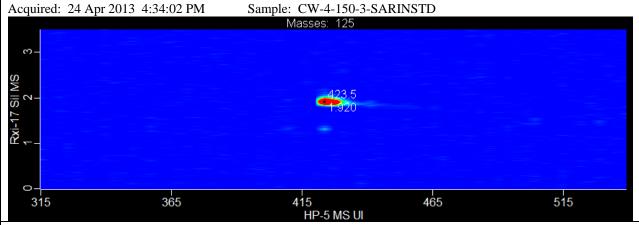
Bottom: Chromatogram of reference chemical of O-Isopropyl methylphosphonofluoridate.



File : C:\LLNL\TOF1335 Method: 2D OPCW020613



File : C:\LLNL\TOF1336 Method: 2D OPCW020613

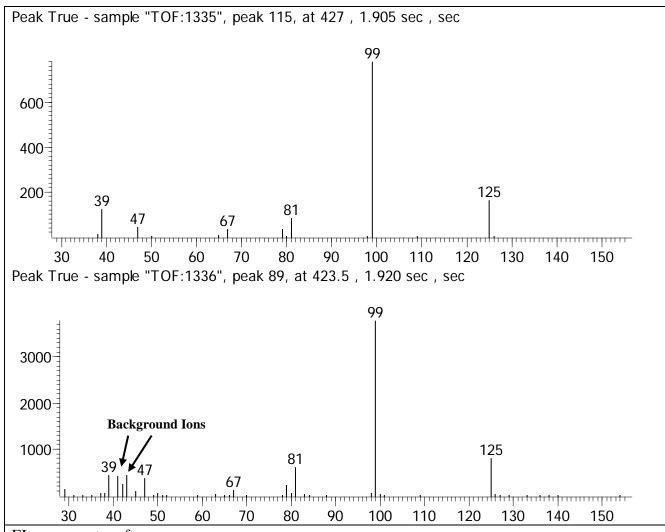


GCxGC EI contour plot supporting identification of Chemical C-1; EIC

Top: Contour plot of the blank.

Center: Contour plot of the sample (aliquot code listed in header).

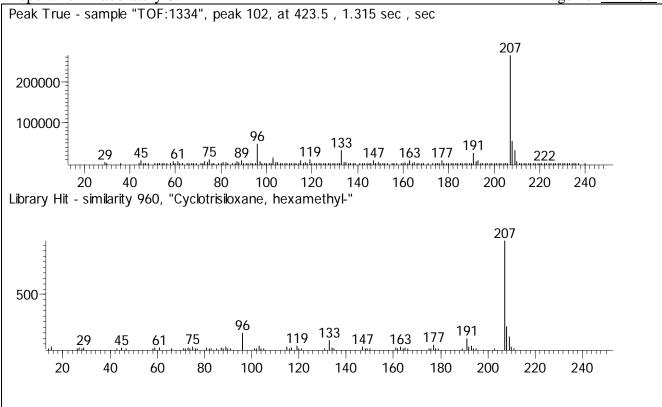
Bottom: Contour plot of reference chemical of O-Isopropyl methylphosphonofluoridate.



EI mass spectra of:

Top: **Chemical C-1** (file name listed in header).

Bottom: Reference chemical of O-Isopropyl methylphosphonofluoridate.



EI mass spectra of:

Top: Background contaminant found in blank within  $\pm$  0.1 min of reported chemical (file name listed in header).

Bottom: Library mass spectrum of hexamethylcyclotrisiloxane.

# **SAMPLE SUMMARY: D**

Sample Code: P-304/07 Laboratory Assigned Code: CW-4-147-4

Description and condition of sample: Approximately 5 mL of plasma

#### Chemical: D-1

Chemical name &	CAS#	Schedule			
O-Isopropyl methylphos					
	107-44-8	1.A.01			
Aliquot(s)	Original/derivative	Analysis technique			
CW-4-150-5-P-304	Original	GC-MS/MS (CI)			
CW-4-150-5-P-304	GC-MS (CI)				
Comments:					

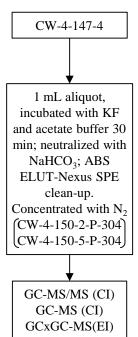
Report from Laboratory 07 Page no. <u>30</u>

# SAMPLE PREPARATION DESCRIPTION: D

1. Sample preparation

Initial Aliquot Code	Type of Sample Preparation	Amount/ Volume	Sample/Blank Preparation Procedures  Added 3 mL of acetate buffer and 220 μL of 4.51 M KF. Incubated 30 minutes at room temperature. Neutralized with 0.8 M Sodium Bicarbonate. ABS ELUT-Nexus cartridge cleanup. Eluted using 2 mL ethyl acetate and reduced sample volume approximately 100 μL.		Resulting Aliquot Code	
CW-4-147-4	Fluoride Reactivation	1 mL			CW-4-150-2-P-304	
CW-4-150-2-P-304	Sample Split	50 μL	Split of sample CW-4-150-2-P-304.	50 μL	CW-4-150-5-P-304	

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#### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by incubating the nerve agent, Sarin, with commercially procured human blood plasma. Cholinesterase activities of the plasma sample were measured using method outline in Ellman (1961) before and after incubation to ensure agent-adduction occurred. The resultant agent-adducted plasma was used for subsequent method development.

Adducted plasma samples were reactivated using the process outlined in Degenhardt *et al* (2004). 1 mL of plasma was added to 3 mL of acetate buffer solution [0.189 M acetic acid; 10.8 mM sodium acetate]. Appropriate amount of Potassium Fluoride [4.51M] was added to achieve a final concentration of 0.25 M KF in the unknown sample work-up. Sample was allowed to incubate at room temperature for 30 minutes. 0.5 mL of 0.8 M sodium bicarbonate was added to neutralize acid.

Several methods were attempted to isolate the reactivated sarin from the sample:

- 4. Direct dichloromethane extraction
- 5. SPE preparation #1

Agilent ABS Elut-NEXUS (200 mg/6 mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

6. SPE preparation #2

Phenomenex Strata-X 33µm (30mg/3mL)

Conditioning: 2 mL N-hexane; 4 mL Ethyl Acetate; 4 mL H<sub>2</sub>O

Load 4.2 mL of reactivated sample.

Elution: 2 mL Ethyl Acetate

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

An Agilent ABS Elut-Nexus SPE cartridge was conditioned, by gravity, with 2 mL of N-hexane, 2x2 mL ethyl acetate, followed by 2x2 mL of  $H_2O$ . 1 mL reactivated plasma sample was then loaded into cartridge and allowed to pass through. Sample was eluted with 2 mL of ethyl acetate into a clean vial. Sample was reduced in volume to approximately 100  $\mu$ L for analysis.

#### References

- G.L. Ellman. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharm 7: 88-95(1961).
- C. E. A. M. Degenhardt, K. Pleijsier, M. J. van der Schans, J. P. Langenberg, K. E. Preston, M. I. Solano, V. L. Maggio, J. R. Barr. Improvements of the Fluoride Reactivation Method for the verification of nerve agent exposure. Journal of Analytical Toxicology. 28(5): 364-371 (2004).

## GC-MS/MS (CI) TECHNIQUE METHOD AND ANALYSIS DESCRIPTION

#### **Identification**

Chemical: D-1

**Aliquot code:** CW-4-150-5-P-304

**Datafile name:** TSQ0334

Compound identified as: Original Compound Compound reference: Reference Chemical (OPCW) Match algorithm and match factor: NIST 948/999

#### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

Column brand/phase: Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 90 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

Solvent delay time: 3 min Electron energy: 100 eV Reaction gas: Ammonia Ionisation polarity: Positive

Scan range/time: 60-200 m/z in 1 second

Mass resolution: 0.7

Type of MS/MS scan: Product ion scan

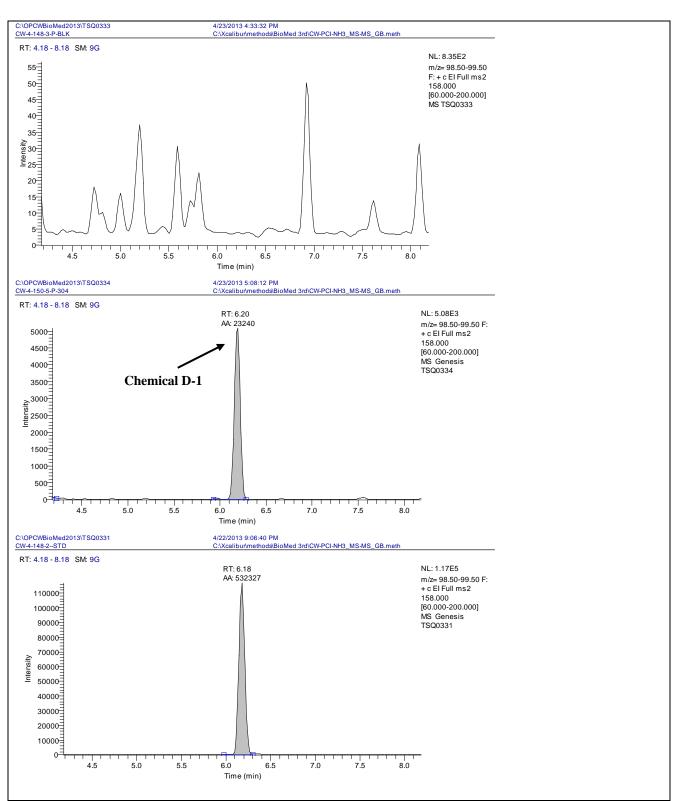
Precursor ion(s): m/z 158 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	Dogult	
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result	
99	23240	1.0	532327	1.0	N/A	N/A	
141	3087	0.133	57494	0.108	±30%	23.0%	

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

#### **Remarks**

<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 

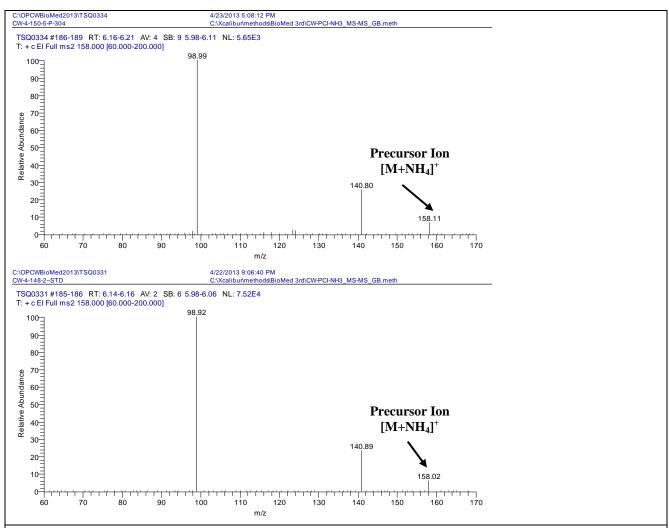


GC-MS/MS (CI) chromatograms supporting identification of Chemical D-1; EIC

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **O-Isopropyl methylphosphonofluoridate.** 



Top: Chemical D-1 (aliquot code listed in header).

Bottom: Reference chemical of O-Isopropyl methylphosphonofluoridate.

### **Identification**

Chemical: D-1

Aliquot code: CW-4-150-5-P-304 Datafile name: CW000630.D

Compound identified as: Original Compound Compound reference: Reference Chemical (OPCW)

### **Analysis Method**

GC Instrument manufacturer and type: Agilent 6890

Carrier gas: Helium

Flow control/rate: Constant Flow, 32 cm/sec Injection mode: Pulsed Splitless, 0.75 min

**Injector temperature:** 250 °C

**Column brand/phase:** Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane **Column Length x ID x Film thickness:** 30 m x 0.25 mm x 0.25 µm

GC temperature programme: 40 °C (3 min), 8 °C/min to 100 °C, 25 °C/min to 300 °C (3 min)

MS Instrument manufacturer and type: Agilent 5975 MSD

**Solvent delay time:** 5.4 min **Electron energy:** 82 eV

Scan range/time: SIM, m/z 141, 158, 100 µs dwell time

Source temperature: 230 °C

Reaction gas: Ammonia and flow: 21%

**Ionisation polarity:** Positive

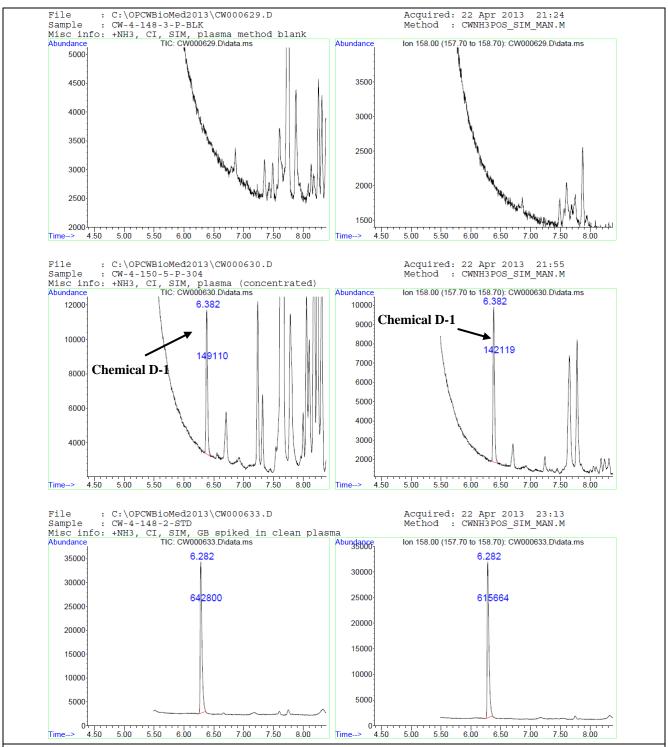
Jana/tuonaitiana		ample	Standard		Criteria	Dogwl4
Ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
141	6306	0.044	28724	0.047	±50%	4.9%
158	142119	1.000	615664	1.000	N/A	N/A

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

### Remarks

Due to the solvent delay of 5.4 min, the chromatogram could not be presented with a window of the retention time of the compound minus 2 min.

<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ; <10%:  $\pm 50\%$ 



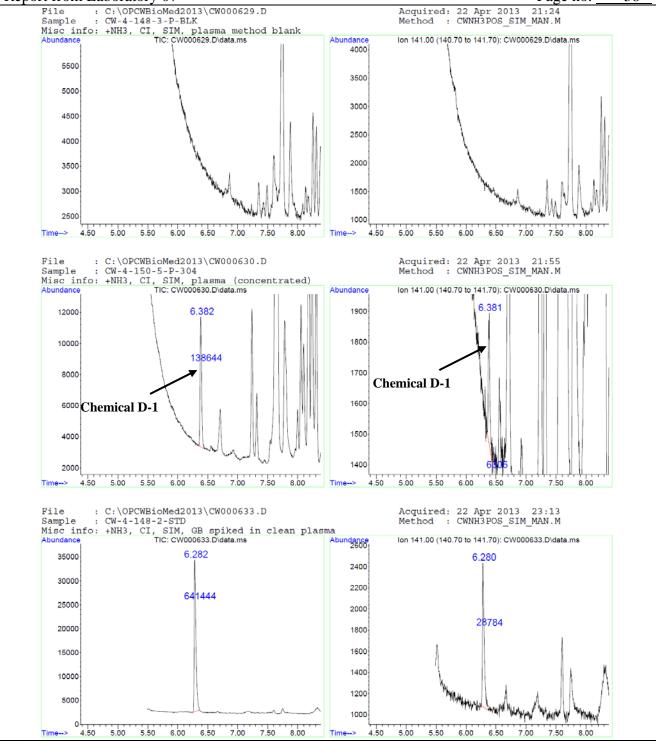
EI chromatograms supporting identification of Chemical D-1;

TIC on left, EIC (m/z 158) on right.

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of O-Isopropyl methylphosphonofluoridate.



EI chromatograms supporting identification of Chemical D-1;

TIC on left, EIC (m/z 141) on right.

Top: Chromatogram of the blank.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of O-Isopropyl methylphosphonofluoridate.

# **SAMPLE SUMMARY: E**

Sample Code: U-305/07 Laboratory Assigned Code: CW-4-147-5

Description and condition of sample: Approximately 10 mL of urine

Chemical: E-1

Chemical name &	Structure	CAS#	Schedule
Isopropyl methylpl	nosphonate		
O OH	1832-54-8	2.B.04	
Aliquot(s)	Original/derivative	Analysis te	chnique
CW-4-151-2-U-305	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
CW-4-151-2-U-305	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
Comments:			

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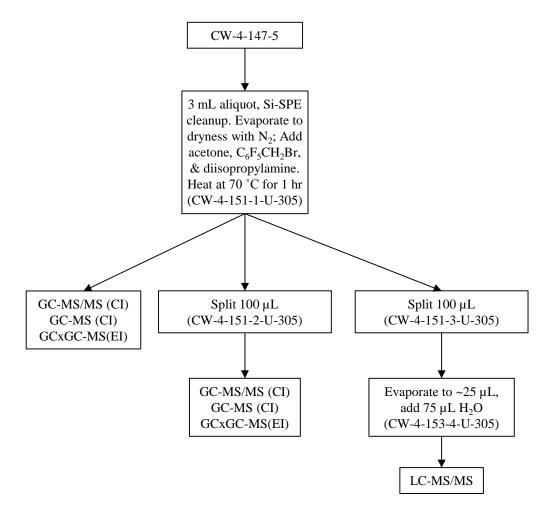
# **SAMPLE PREPARATION DESCRIPTION: E**

1. Sample preparation

Initial Aliquot Code	Type of Sample Preparation	Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code
CW-4-147-5	Urine Clean-Up and Pentafluorobenzyl Derivatization	3 mL	Silica SPE cartridge cleanup. Eluted using 3 mL 25% H <sub>2</sub> O in acetonitrile. Reduced sample volume to complete dryness with nitrogen gas. Added 300 μL of acetone, 5 μL pentafluorobenzyl bromide and 5 μL diisopropylamine. Sample heated at 70°C for one hour.	310 μL	CW-4-151-1-U-30:
CW-4-151-1-U-305	Sample Split	100 μL	Split of sample CW-4-151-1-U-305.	100 μL	CW-4-151-2-U-30
CW-4-151-1-U-305	Sample Split	100 μL	Split of sample CW-4-151-1-U-305.	100 μL	CW-4-151-3-U-305
CW-4-151-3-U-305	Preparation for LC-MS Analysis	100 μL	Reduced volume to ~25 $\mu L$ with nitrogen gas. Added 75 $\mu L$ of $H_2O$ .	100 μL	CW-4-153-4-U-305

## 2. Additional information

SPE=solid phase extraction



### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by spiking a mixture of methylphosphonic acid, ethyl methylphosphonic acid, isopropyl methylphosphonic acid, and pinacolyl methylphosphonic acid into commercially procured human urine. The resultant acid-containing urine was used for subsequent method development.

Several SPE methods were attempted to isolate the phosphonic acids from the urine sample:

1. Alltech Silica (500 mg/4 mL)

Conditioning: 25% H<sub>2</sub>O in acetonitrile; 3 mL of acetonitrile

Load 3x1 mL of urine sample.

Wash: 2mL acetonitrile; 2 mL 10% H<sub>2</sub>O in acetonitrile.

Elution: 25% H<sub>2</sub>O (in acetonitrile)

2. Agilent ABS Elut-NEXUS (200 mg/ 6 mL)

Conditioning: N-hexane; Ethyl Acetate; H<sub>2</sub>O

Load 3x1 mL of urine sample

Elution: Ethyl Acetate

3. Phenomenex Strata-X 33µm (30 mg/3 mL)

Sample pretreatment: Dilute sample 1:1 with Acetate Buffer (pH ~3.5)

Conditioning: Methanol; Acetate Buffer (pH~3.5)

Load: 1 mL of urine sample

Wash: Acetate Buffer (pH ~3.5); methanol Elution: 5% ammonium hydroxide in methanol

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

Spiked urine samples were processed following cleanup conditions found in Mawhinney (2007). An Alltech silica SPE was conditioned with 4mL 25% water in acetonitrile, followed by 3 mL of acetonitrile. Sample, 3x1 mL, was loaded onto cartridge and washed with 2 mL acetonitrile and 2 mL of 10% water in acetonitrile. Samples were eluted with using 25% water in acetonitrile.

Mawhinney took urine sample to dryness and reconstituted before introduction to SPE cartridge; however, in a side by side comparison, we found loading urine directly onto a conditioned cartridge resulted in the best recovery. Additionally, success has been reported with using polymeric SPE for cleanup and although we found success with one polymeric SPE, our best recovery was using silica SPE.

The urine sample eluted from the SPE cartridge was derivatized before analysis. The eluted sample was taken to dryness. Following the process outlined in Palit (2004), the dried samples were derivatized by adding 5  $\mu$ L pentafluorobenzyl bromide, 5  $\mu$ L diisopropylamine and 300  $\mu$ L acetone and heating to 70°C for 1 hour.

### References

Mawhinney, D.B., Hameli, E.I., Fraser, R., Silva, S.S., Pavlopoulos, A.J. Kobelski, R.J. J.; The determination of organophosphate nerve agent metabolites in human urine by hydrophilic interaction liquid chromatography tandem mass spectrometry. J. Chromatogr. B. 852 (2007) 235-243.

Palit, M., Gupta, A.K., Jain, R. and Raza, S.K.; Determination of pentafluorobenzyl derivatives of phosphonic and phosphonothionic acids by gas chromatography-mass spectrometry. J Chromatogr. A, 1043 (2004) 275-284.

### **Identification**

**Chemical:** E-1

**Aliquot code:** CW-4-151-2-U-305

**Datafile name:** TSQ0358

**Compound identified as:** Pentafluorobenzyl derivative **Compound reference:** Reference Chemical (own synthesis)

Match algorithm and match factor: NIST, 981/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

Column brand/phase: Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness:  $30 \text{ m} \times 0.25 \text{ mm} \times 0.25 \text{ } \mu\text{m}$ 

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

**Solvent delay time:** 3 min **Electron energy:** 100 eV **Reaction gas:** Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

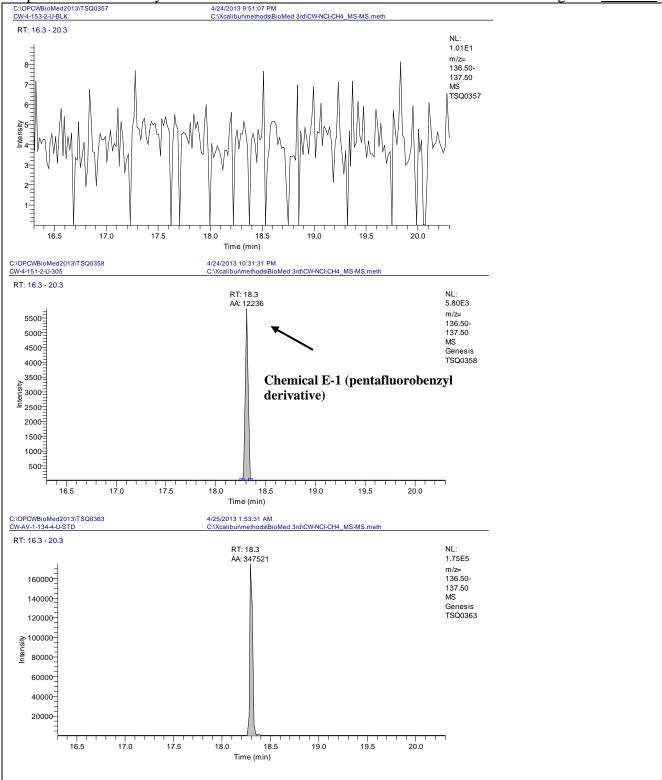
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 137 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	Dogult
ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	6684	0.104	243159	0.109	±30%	4.4%
95	61457	1.0	2338092	1.0	N/A	N/A
137	12236	0.149	347521	0.199	±30%	33.0%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

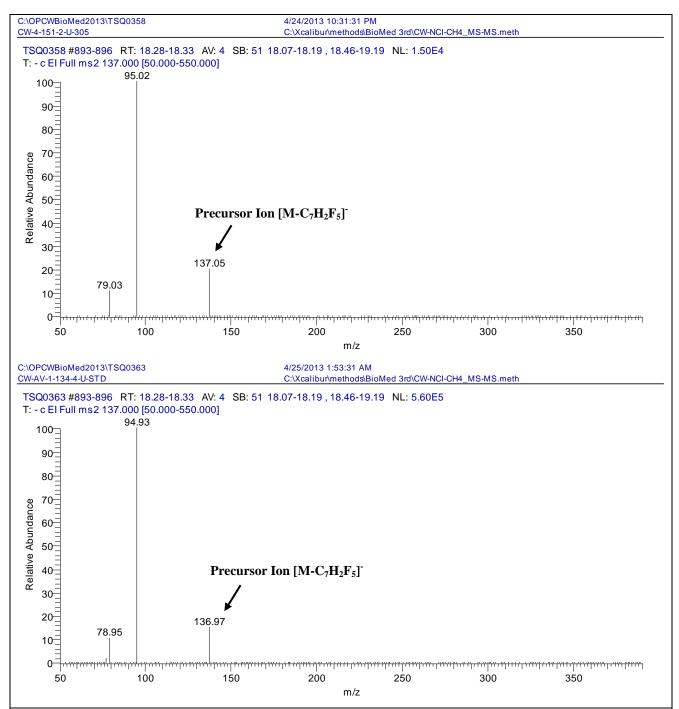
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical E-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of isopropyl methylphosphonate.** 



Top: Chemical E-1 (pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of isopropyl methylphosphonate.

### **Identification**

**Chemical:** E-1

**Aliquot code:** CW-4-151-2-U-305

**Datafile name:** TSQ0379

**Compound identified as:** Pentafluorobenzyl derivative **Compound reference:** Reference Chemical (own synthesis)

Match algorithm and match factor: NIST, 848/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

**Column brand/phase:** Agilent DB-1: 100% Dimethylpolysiloxane **Column Length x ID x Film thickness:** 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

**Solvent delay time:** 3 min **Electron energy:** 100 eV **Reaction gas:** Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

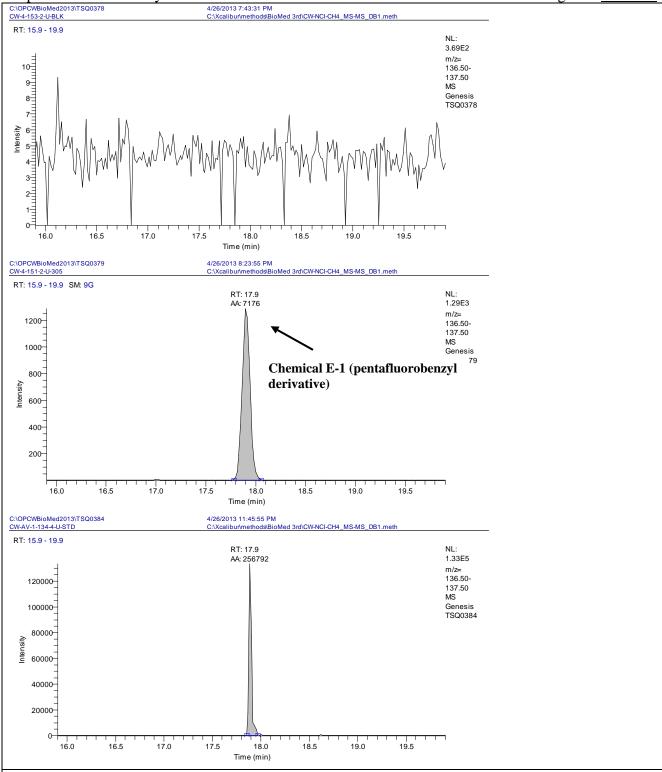
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 137 Collision gas: Argon Collision Energy: 10

Tomaltmanaiti and		ample	Standard		Criteria	D carel4	
Ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result	
79	2700	0.022	211094	0.113	±30%	78.1%	
95	120425	1.000	1862500	1.000	N/A	N/A	
137	7176	0.109	256792	0.138	±30%	21.0%	

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

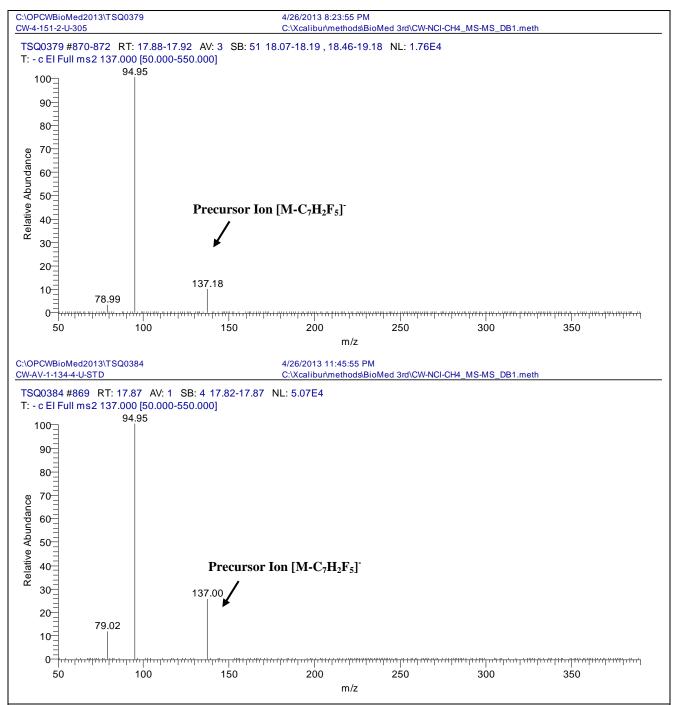
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\leq 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical E-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of isopropyl methylphosphonate.** 



Top: Chemical E-1 (pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of isopropyl methylphosphonate.

# **SAMPLE SUMMARY: F**

Sample Code: U-306/07 Laboratory Assigned Code: CW-4-147-6

Description and condition of sample: Approximately 10 mL of urine

## **Chemical:** F-1

Chemical name &	CAS#	Schedule	
Isopropyl methylpl	nosphonate		
OH OH	1832-54-8	2.B.04	
Aliquot(s)	Original/derivative	Analysis te	chnique
CW-4-152-2-U-306	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
CW-4-152-2-U-306	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
Comments:			

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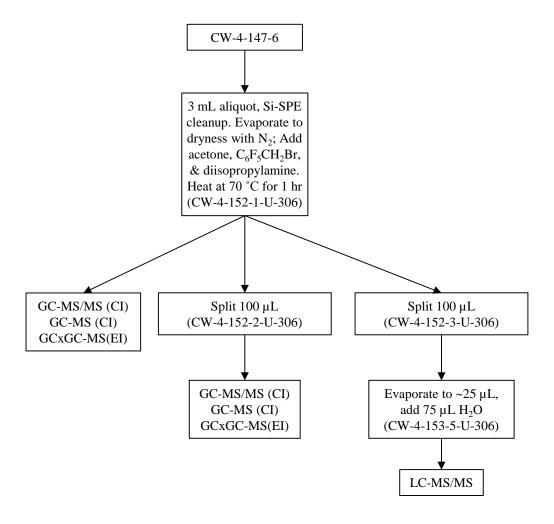
# **SAMPLE PREPARATION DESCRIPTION: F**

1. Sample preparation

Initial Aliquot Code	Type of Sample Preparation	Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code
CW-4-147-6	Urine Clean-Up and Pentafluorobenzyl Derivatization	3 mL	Silica SPE cartridge cleanup. Eluted using 3 mL 25% H <sub>2</sub> O in acetonitrile. Reduced sample volume to complete dryness with nitrogen gas. Added 300 μL of acetone, 5 μL pentafluorobenzyl bromide and 5 μL diisopropylamine. Sample heated at 70°C for one hour.	310 μL	CW-4-152-1-U-306
CW-4-152-1-U-306	Sample Split	100 μL	Split of sample CW-4-152-1-U-306.	100 μL	CW-4-152-2-U-300
CW-4-152-1-U-306	Sample Split	100 μL	Split of sample CW-4-152-1-U-306.	100 μL	CW-4-152-3-U-306
CW-4-152-3-U-306	Preparation for LC-MS Analysis	100 μL	Reduced volume to ~25 $\mu L$ with nitrogen gas. Added 75 $\mu L$ of $H_2O$ .	100 μL	CW-4-153-5-U-306

## 2. Additional information

SPE=solid phase extraction



### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by spiking a mixture of methylphosphonic acid, ethyl methylphosphonic acid, isopropyl methylphosphonic acid, and pinacolyl methylphosphonic acid into commercially procured human urine. The resultant acid-containing urine was used for subsequent method development.

Several SPE methods were attempted to isolate the phosphonic acids from the urine sample:

1. Alltech Silica (500 mg/4 mL)

Conditioning: 25% H<sub>2</sub>O in acetonitrile; 3 mL of acetonitrile

Load 3x1 mL of urine sample.

Wash: 2mL acetonitrile; 2 mL 10% H<sub>2</sub>O in acetonitrile.

Elution: 25% H<sub>2</sub>O (in acetonitrile)

2. Agilent ABS Elut-NEXUS (200 mg/ 6 mL)

Conditioning: N-hexane; Ethyl Acetate; H<sub>2</sub>O

Load 3x1 mL of urine sample

Elution: Ethyl Acetate

3. Phenomenex Strata-X 33µm (30 mg/3 mL)

Sample pretreatment: Dilute sample 1:1 with Acetate Buffer (pH ~3.5)

Conditioning: Methanol; Acetate Buffer (pH~3.5)

Load: 1 mL of urine sample

Wash: Acetate Buffer (pH ~3.5); methanol Elution: 5% ammonium hydroxide in methanol

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

Spiked urine samples were processed following cleanup conditions found in Mawhinney (2007). An Alltech silica SPE was conditioned with 4mL 25% water in acetonitrile, followed by 3 mL of acetonitrile. Sample, 3x1 mL, was loaded onto cartridge and washed with 2 mL acetonitrile and 2 mL of 10% water in acetonitrile. Samples were eluted with using 25% water in acetonitrile.

Mawhinney took urine sample to dryness and reconstituted before introduction to SPE cartridge; however, in a side by side comparison, we found loading urine directly onto a conditioned cartridge resulted in the best recovery. Additionally, success has been reported with using polymeric SPE for cleanup and although we found success with one polymeric SPE, our best recovery was using silica SPE.

The urine sample eluted from the SPE cartridge was derivatized before analysis. The eluted sample was taken to dryness. Following the process outlined in Palit (2004), the dried samples were derivatized by adding 5  $\mu$ L pentafluorobenzyl bromide, 5  $\mu$ L diisopropylamine and 300  $\mu$ L acetone and heating to 70°C for 1 hour.

### References

Mawhinney, D.B., Hameli, E.I., Fraser, R., Silva, S.S., Pavlopoulos, A.J. Kobelski, R.J. J.; The determination of organophosphate nerve agent metabolites in human urine by hydrophilic interaction liquid chromatography tandem mass spectrometry. J. Chromatogr. B. 852 (2007) 235-243.

Palit, M., Gupta, A.K., Jain, R. and Raza, S.K.; Determination of pentafluorobenzyl derivatives of phosphonic and phosphonothionic acids by gas chromatography-mass spectrometry. J Chromatogr. A, 1043 (2004) 275-284.

### Identification

Chemical: F-1

**Aliquot code:** CW-4-152-2-U-306

**Datafile name:** TSQ0360

**Compound identified as:** Pentafluorobenzyl derivative **Compound reference:** Reference Chemical (own synthesis)

Match algorithm and match factor: NIST, 920/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

Column brand/phase: Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

**Solvent delay time:** 3 min **Electron energy:** 100 eV **Reaction gas:** Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

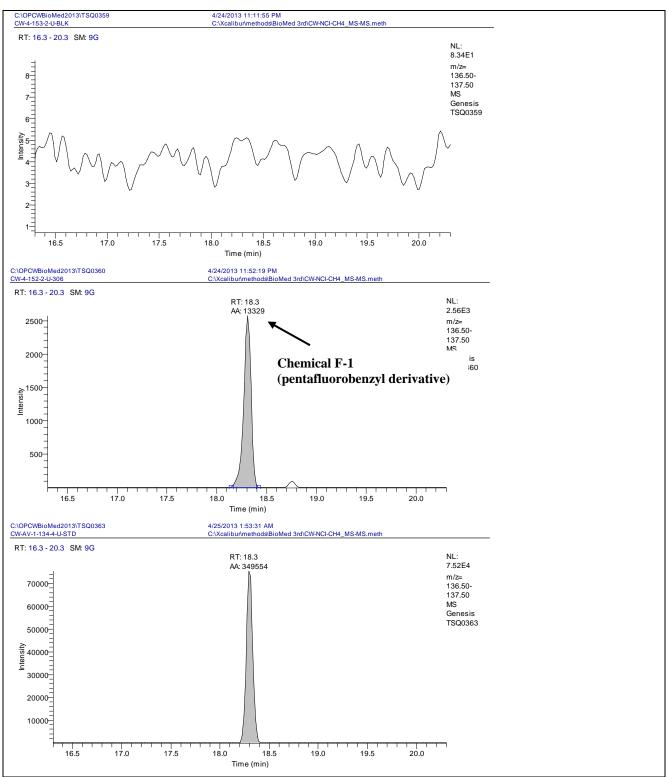
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 137 Collision gas: Argon Collision Energy: 10

Iong/twongitions	Sample		Standard		Criteria	Dogult
Ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	3424	0.025	243296	0.104	±30%	73.2%
95	139394	1	2338092	1.0	N/A	N/A
137	13329	0.096	347521	0.199	±30%	27.2%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

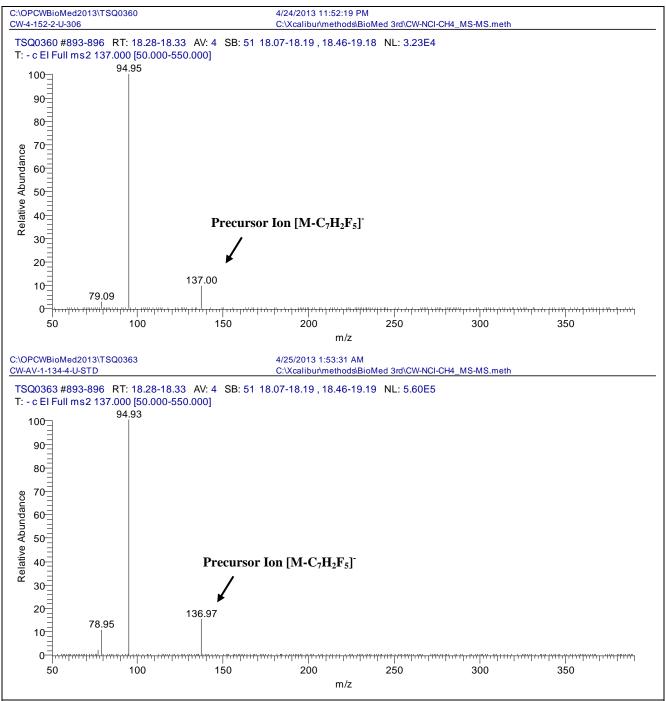
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical F-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of isopropyl methylphosphonate.** 



Top: Chemical F-1(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of **Pentafluorobenzyl derivative of isopropyl methylphosphonate.** 

### **Identification**

**Chemical:** F-1

**Aliquot code:** CW-4-152-2-U-306

**Datafile name:** TSQ0381

Compound identified as: Pentafluorobenzyl derivative Compound reference: Reference Chemical (own synthesis) Match algorithm and match factor: NIST, 820/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

**Column brand/phase:** Agilent DB-1: 100% Dimethylpolysiloxane **Column Length x ID x Film thickness:** 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

Solvent delay time: 3 min Electron energy: 100 eV Reaction gas: Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

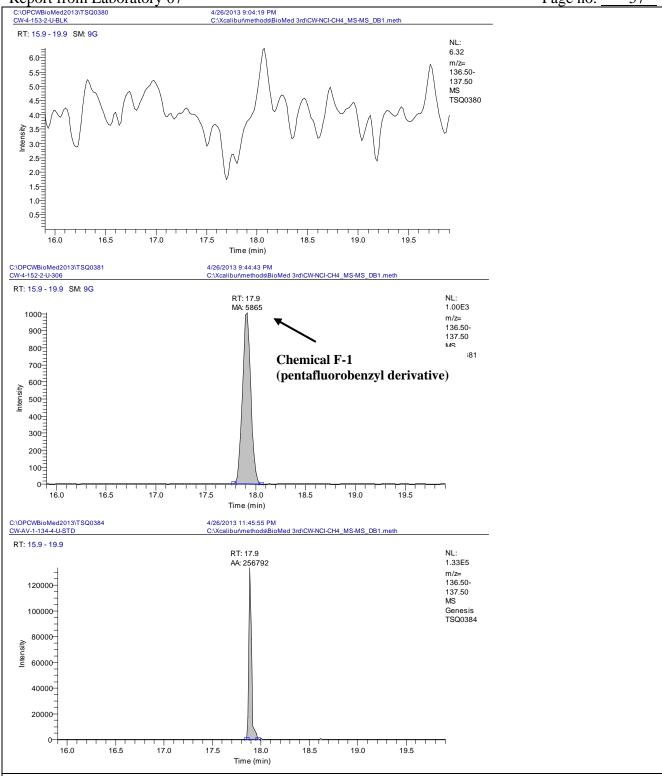
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 137 Collision gas: Argon Collision Energy: 10

T //	Sa	ample	Standard		Criteria	D 1/
Ions/transitions	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	2700	0.022	211094	0.113	±30%	80.2%
95	120425	1.000	1862500	1.000	N/A	N/A
137	5865	0.049	256792	0.138	±30%	64.7%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

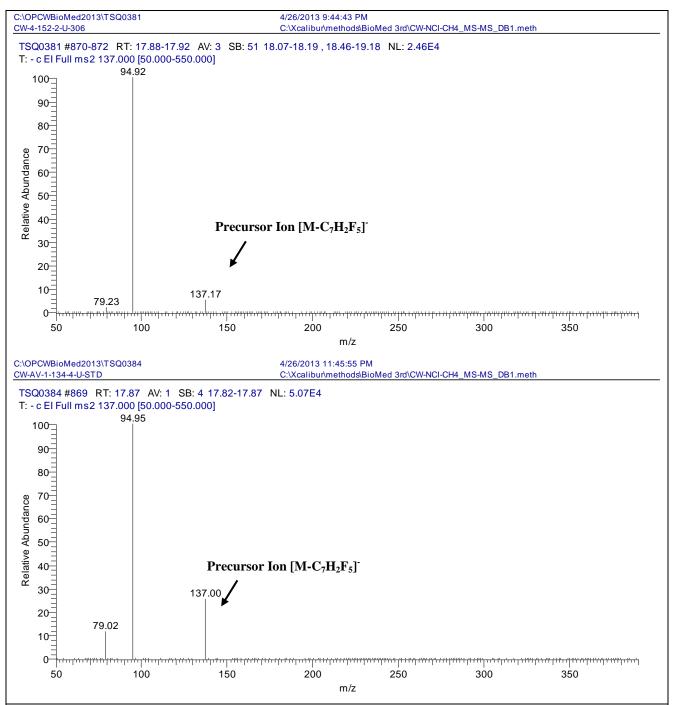
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm20\%$ ; >20% to 50%:  $\pm25\%$ ; >10% to 20%:  $\pm30\%$ ;  $\leq10\%$ :  $\pm50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical F-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of isopropyl methylphosphonate.** 



Top: Chemical F-1(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of isopropyl methylphosphonate.

# **SAMPLE SUMMARY: G**

Sample Code: U-307/07 Laboratory Assigned Code: CW-4-147-7

Description and condition of sample: Approximately 10 mL of urine

## Chemical: G-1

Chemical name &	Structure	CAS#	Schedule
Ethyl methylpho	sphonate		
OH OH	1832-53-7	2.B.04	
Aliquot(s)	Original/derivative	Analysis te	chnique
CW-4-152-5-U-307	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
CW-4-152-5-U-307	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
Comments:			•

### Chemical: G-2

Chemical name	& Structure	CAS#	Schedule
Pinacolyl methy	lphosphonate		
HO HO		616-52-4	2.B.04
Aliquot(s)	Original/derivative	Analysis te	chnique
CW-4-152-5-U-307	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
CW-4-152-5-U-307	Pentafluorobenzyl derivative	GC-MS/N	IS(CI)
CW-4-153-6-U-307	Pentafluorobenzyl derivative	LC-MS	/MS
Comments:			

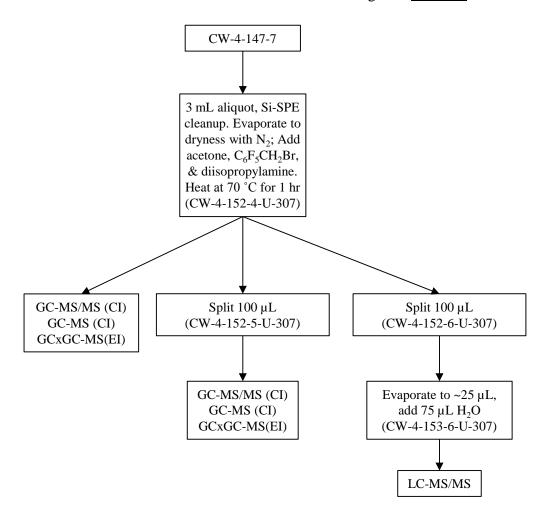
# **SAMPLE PREPARATION DESCRIPTION: G**

1. Sample preparation

Initial Aliquot Code	Type of Sample Preparation	Amount/ Volume	Sample/Blank Preparation Procedures	End Volume	Resulting Aliquot Code
CW-4-147-7	Urine Clean-Up and Pentafluorobenzyl Derivatization	3 mL	Silica SPE cartridge cleanup. Eluted using 3 mL 25% H <sub>2</sub> O in acetonitrile. Reduced sample volume to complete dryness with nitrogen gas. Added 300 μL of acetone, 5 μL pentafluorobenzyl bromide and 5 μL diisopropylamine. Sample heated at 70°C for one hour.	310 μL	CW-4-152-4-U-307
CW-4-152-4-U-307	Sample Split	100 μL	Split of sample CW-4-152-4-U-307.	100 μL	CW-4-152-5-U-307
CW-4-152-4-U-307	Sample Split	100 μL	Split of sample CW-4-152-4-U-307.	100 μL	CW-4-152-6-U-307
CW-4-152-6-U-307	Preparation for LC-MS Analysis	100 μL	Reduced volume to ~25 $\mu L$ with nitrogen gas. Added 75 $\mu L$ of $H_2O$ .	100 μL	CW-4-153-6-U-307

## 2. Additional information

SPE=solid phase extraction



### Description of sample preparation and analysis methods

Sample preparation and analysis methods were developed using an in-house standard made by spiking a mixture of methylphosphonic acid, ethyl methylphosphonic acid, isopropyl methylphosphonic acid, and pinacolyl methylphosphonic acid into commercially procured human urine. The resultant acid-containing urine was used for subsequent method development.

Several SPE methods were attempted to isolate the phosphonic acids from the urine sample:

1. Alltech Silica (500 mg/4 mL)

Conditioning: 25% H<sub>2</sub>O in acetonitrile; 3 mL of acetonitrile

Load 3x1 mL of urine sample.

Wash: 2mL acetonitrile; 2 mL 10% H<sub>2</sub>O in acetonitrile.

Elution: 25% H<sub>2</sub>O (in acetonitrile)

2. Agilent ABS Elut-NEXUS (200 mg/ 6 mL)

Conditioning: N-hexane; Ethyl Acetate; H<sub>2</sub>O

Load 3x1 mL of urine sample

Elution: Ethyl Acetate

3. Phenomenex Strata-X 33µm (30 mg/3 mL)

Sample pretreatment: Dilute sample 1:1 with Acetate Buffer (pH ~3.5)

Conditioning: Methanol; Acetate Buffer (pH~3.5)

Load: 1 mL of urine sample

Wash: Acetate Buffer (pH ~3.5); methanol Elution: 5% ammonium hydroxide in methanol

The following procedure describes, in detail, the method that was determined to be most successful and used for the sample (SPE preparation #1):

Spiked urine samples were processed following cleanup conditions found in Mawhinney (2007). An Alltech silica SPE was conditioned with 4mL 25% water in acetonitrile, followed by 3 mL of acetonitrile. Sample, 3x1 mL, was loaded onto cartridge and washed with 2 mL acetonitrile and 2 mL of 10% water in acetonitrile. Samples were eluted with using 25% water in acetonitrile.

Mawhinney took urine sample to dryness and reconstituted before introduction to SPE cartridge; however, in a side by side comparison, we found loading urine directly onto a conditioned cartridge resulted in the best recovery. Additionally, success has been reported with using polymeric SPE for cleanup and although we found success with one polymeric SPE, our best recovery was using silica SPE.

The urine sample eluted from the SPE cartridge was derivatized before analysis. The eluted sample was taken to dryness. Following the process outlined in Palit (2004), the dried samples were derivatized by adding 5  $\mu$ L pentafluorobenzyl bromide, 5  $\mu$ L diisopropylamine and 300  $\mu$ L acetone and heating to 70°C for 1 hour.

### References

Mawhinney, D.B., Hameli, E.I., Fraser, R., Silva, S.S., Pavlopoulos, A.J. Kobelski, R.J. J.; The determination of organophosphate nerve agent metabolites in human urine by hydrophilic interaction liquid chromatography tandem mass spectrometry. J. Chromatogr. B. 852 (2007) 235-243.

Palit, M., Gupta, A.K., Jain, R. and Raza, S.K.; Determination of pentafluorobenzyl derivatives of phosphonic and phosphonothionic acids by gas chromatography-mass spectrometry. J Chromatogr. A, 1043 (2004) 275-284.

### Identification

Chemical: G-1

**Aliquot code:** CW-4-152-5-U-307

**Datafile name:** TSQ0362

**Compound identified as:** Pentafluorobenzyl derivative **Compound reference:** Reference Chemical (own synthesis)

Match algorithm and match factor: NIST, 804/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

**Column brand/phase:** Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

**Solvent delay time:** 3 min **Electron energy:** 100 eV **Reaction gas:** Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

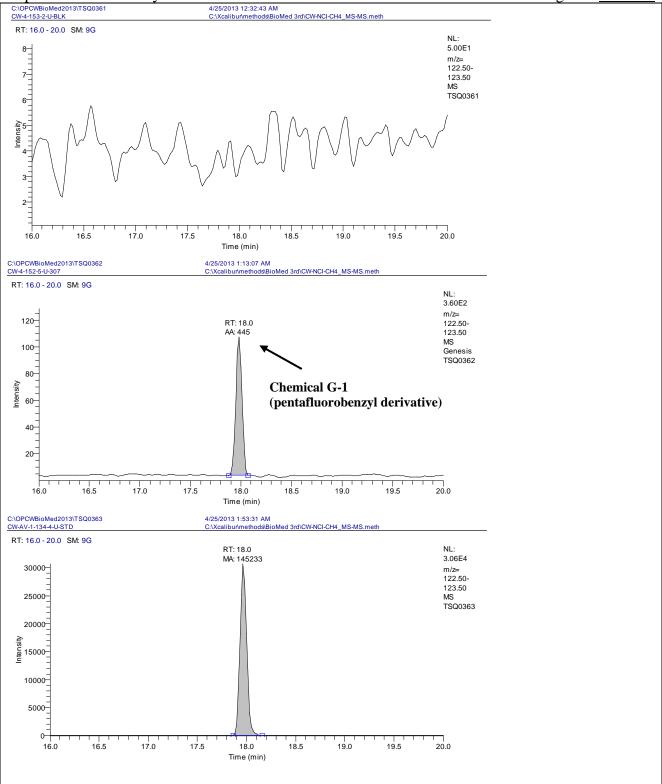
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 123 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	Dogult
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	2358	0.049	307914	0.178	±30%	72.4%
95	48048	1	1731474	1	N/A	
123	445	0.009	145233	0.084	±50%	89.0%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

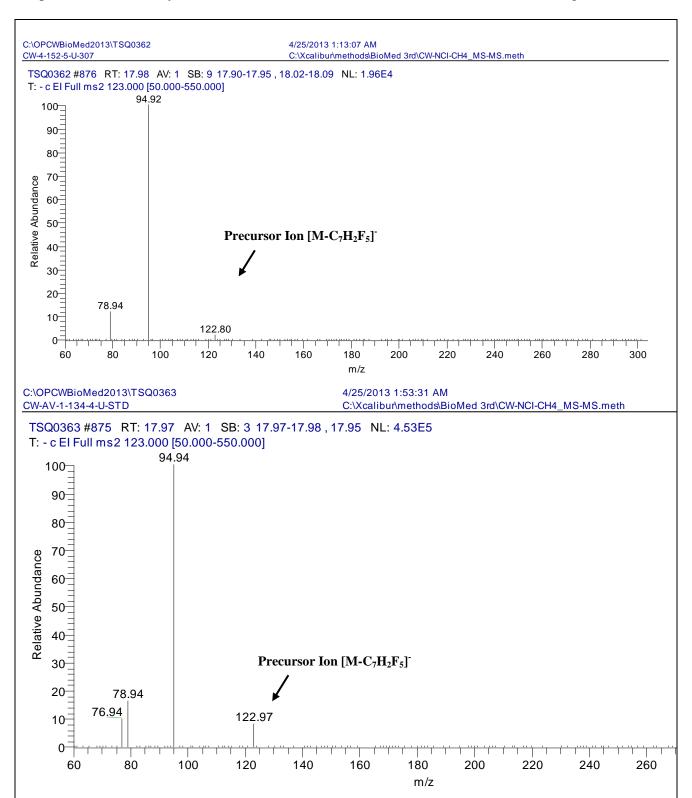
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical G-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of ethyl methylphosphonate.** 



Top: Chemical G-1(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of ethyl methylphosphonate.

### **Identification**

Chemical: G-1

**Aliquot code:** CW-4-152-5-U-307

**Datafile name:** TSQ0383

Compound identified as: Pentafluorobenzyl derivative Compound reference: Reference Chemical (own synthesis) Match algorithm and match factor: NIST, 851/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

Column brand/phase: Agilent DB-1: 100% Dimethylpolysiloxane Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

**Solvent delay time:** 3 min **Electron energy:** 100 eV **Reaction gas:** Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

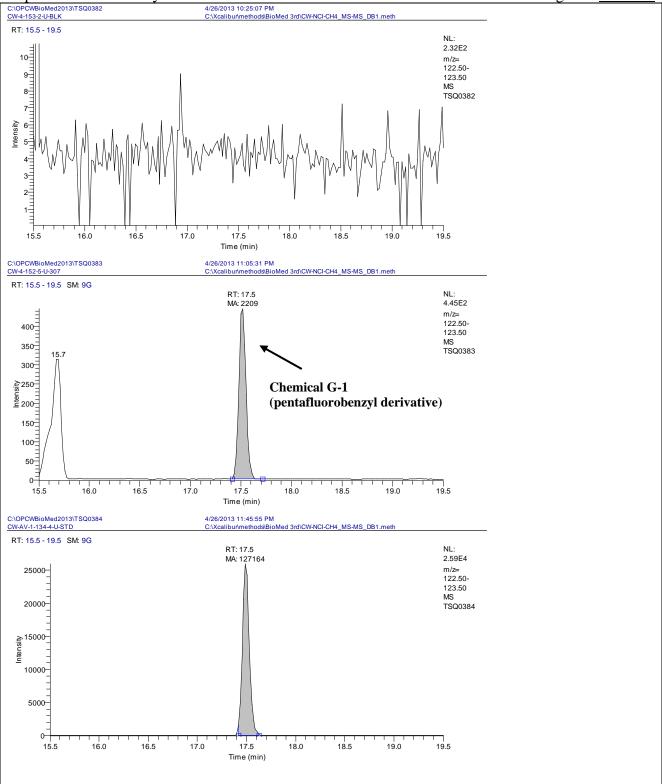
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 123 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	D 14
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	2141	0.038	293708	0.182	±30%	79.4%
95	57029	1.000	1610485	1.000	N/A	N/A
123	2209	0.039	127164	0.079	±50%	40.9%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

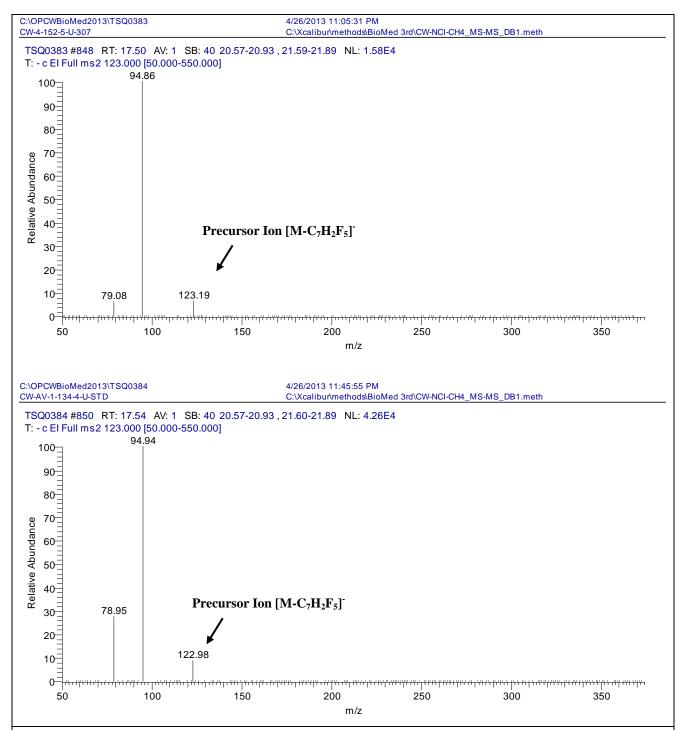
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical G-1** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of ethyl methylphosphonate.** 



Top: Chemical G-1(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of ethyl methylphosphonate.

### **Identification**

Chemical: G-2

**Aliquot code:** CW-4-152-5-U-307

**Datafile name:** TSQ0362

Compound identified as: Pentafluorobenzyl derivative Compound reference: Reference Chemical (own synthesis) Match algorithm and match factor: NIST, 815/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

Column brand/phase: Agilent HP-5MS: (5%-Phenyl)-methylpolysiloxane

Column Length x ID x Film thickness: 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

Solvent delay time: 3 min Electron energy: 100 eV Reaction gas: Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

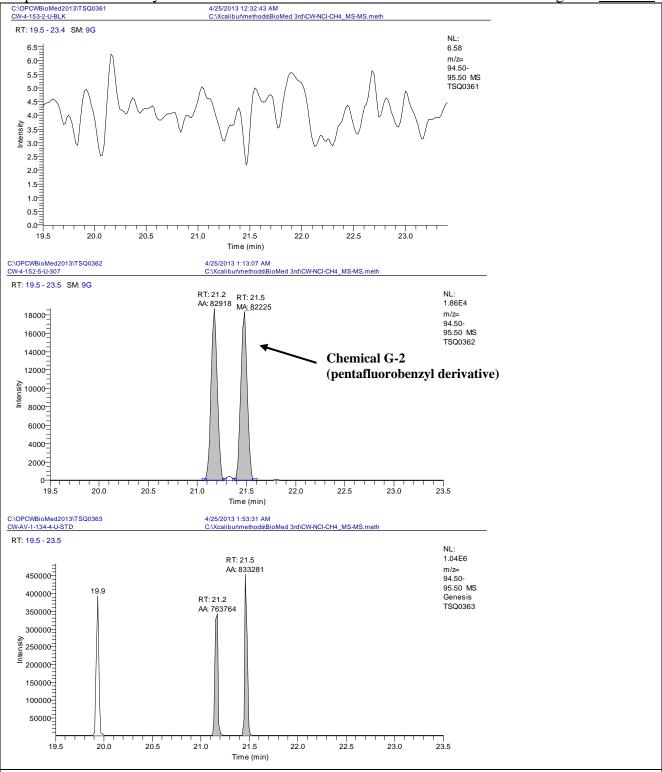
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 179 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	D 14
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance#	Result
79	14388	0.175	128937	0.155	±30%	13.1%
95	82225	1.0	833281	1.0	N/A	N/A
179	5910	0.067	60141	0.072	±50%	7.0%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

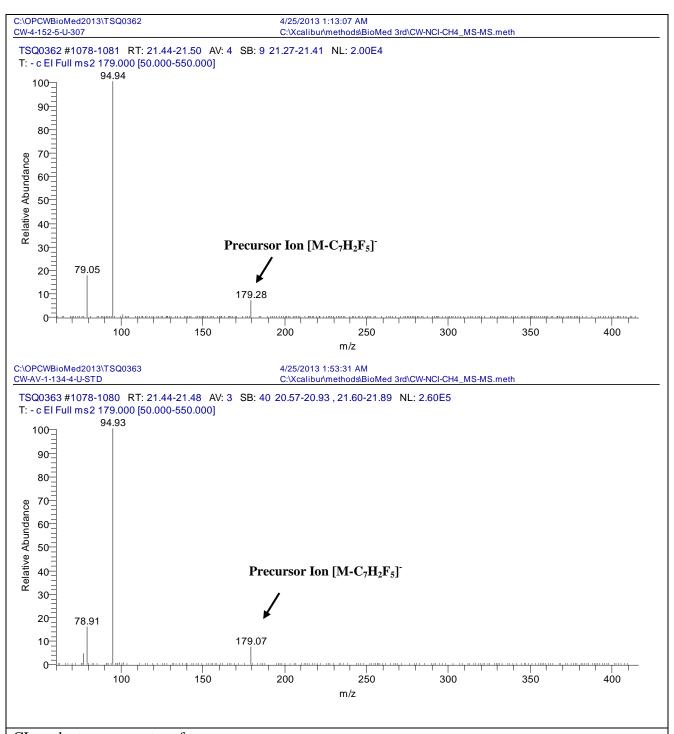
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\le 10\%$ :  $\pm 50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical G-2** (pentafluorobenzyl derivative); TIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of pinacolyl methylphosphonate.** 



Top: Chemical G-2(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of pinacolyl methylphosphonate.

### **Identification**

Chemical: G-2

**Aliquot code:** CW-4-152-5-U-307

**Datafile name:** TSQ0383

Compound identified as: Pentafluorobenzyl derivative Compound reference: Reference Chemical (own synthesis) Match algorithm and match factor: NIST, 931/999

### **Analysis Method**

GC Instrument manufacturer and type: Thermo Fisher Trace GC Ultra

Carrier gas: Helium

Flow control/rate: Constant flow, 1 mL/min

**Injection mode:** Splitless, 0.60 min **Injector temperature:** 230 °C

**Column brand/phase:** Agilent DB-1: 100% Dimethylpolysiloxane **Column Length x ID x Film thickness:** 30 m x 0.25 mm x 0.25 mm

GC temperature programme: 40 °C (3 min), 8 °C/min to 215 °C, 20 °C/min to 300 °C (1 min)

MS Instrument manufacturer and type: Thermo Fisher TSQ Quantum

Solvent delay time: 3 min Electron energy: 100 eV Reaction gas: Methane

**Ionisation polarity:** Negative

Scan range/time: 50-500 m/z in 1 second

Mass resolution: 0.7

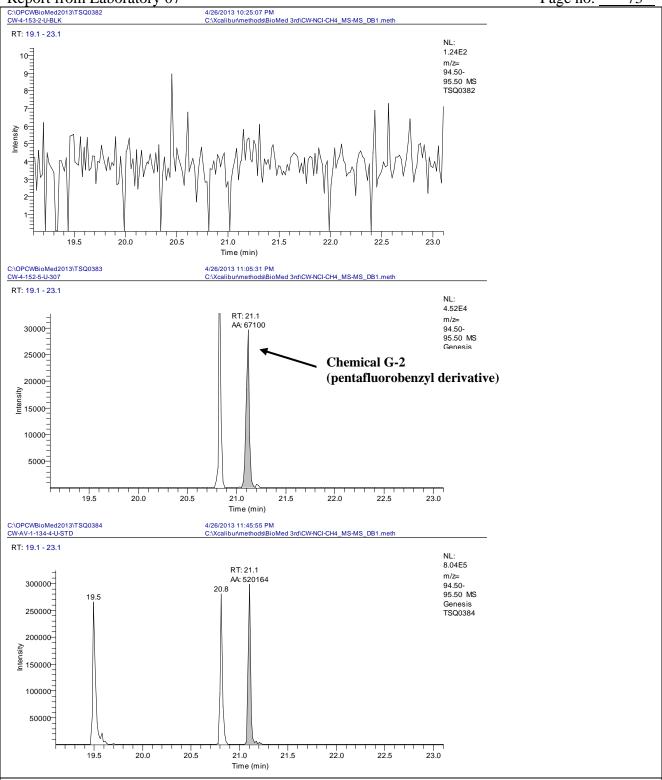
Type of MS/MS scan: Product ion scan

Precursor ion(s): m/z 179 Collision gas: Argon Collision Energy: 10

Ions/transitions	Sample		Standard		Criteria	D14
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
79	6607	0.098	79743	0.153	±30	35.8%
95	67100	1.000	520164	1.000	N/A	N/A
179	3430	0.051	37110	0.071	±50%	28.3%

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

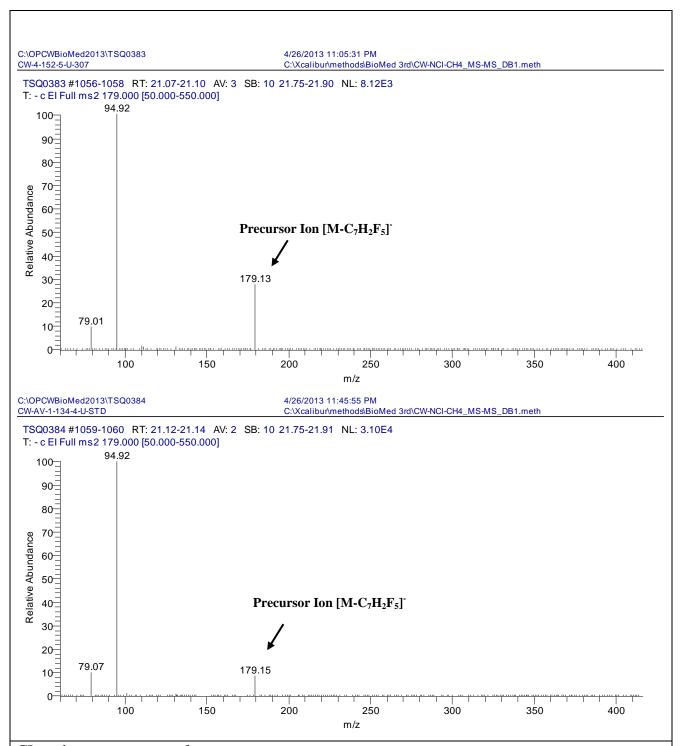
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm20\%$ ; >20% to 50%:  $\pm25\%$ ; >10% to 20%:  $\pm30\%$ ;  $\leq10\%$ :  $\pm50\%$ 



GC-MS/MS (CI) chromatograms supporting identification of **Chemical G-2** (pentafluorobenzyl derivative); EIC

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of pinacolyl methylphosphonate.** 



Top: Chemical G-2(pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of pinacolyl methylphosphonate.

## LC-API-MS/MS TECHNIQUE METHOD AND ANALYSIS DESCRIPTION

### **Identification**

Chemical: G-2

Aliquot code: CW-4-153-6-U-307 Datafile name: ORBI004531

**Compound identified as:** Pentafluorobenzyl Derivative **Compound reference:** Reference Chemical (own synthesis)

### **Analysis Method**

**LC Instrument manufacturer and type:** Thermo Surveyor Plus

Injection volume: 10 µL

**Eluent composition:**  $A = H_2O$  with 0.1% formic acid

B = acetonitrile with 0.1% formic acid

**Elution programme:** 95% A for 5 min, linear gradient to 20% A in 10 min, hold at 20% A for 10

min, regenerate column by returning to 95% A in 3 min, hold at 95% A for

10 min.

Flow rate: 200 µL/min

Column brand/phase: Waters Atlantis T3/C18

Column Length x ID x Particle size: 150 mm x 2.1 mm x 3 µm

Column temperature: 30 °C

MS Instrument manufacturer and type: Thermo LTQ XL

**Ionization type:** Electrospray **Ionisation polarity:** Positive

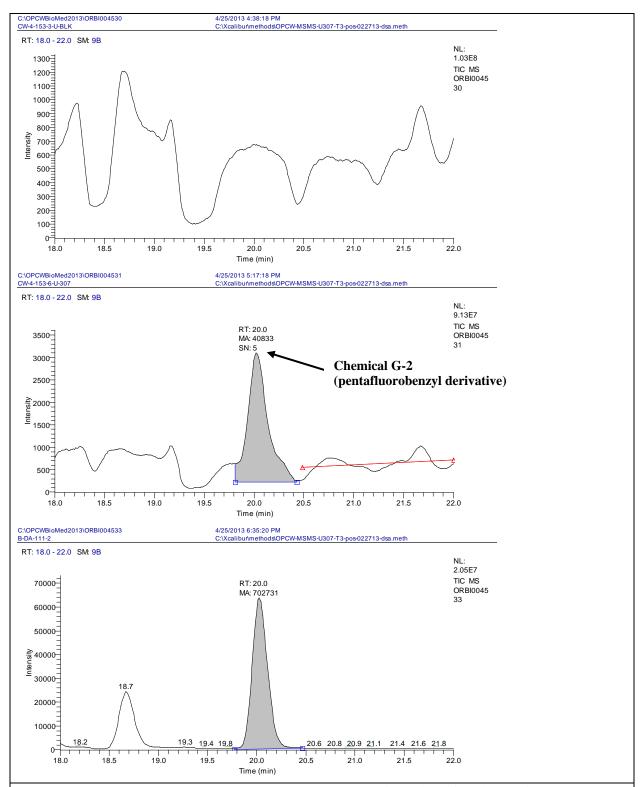
**Electrospray/APCI voltage:** 4.0 kV **Type of MS/MS scan**: Product ion scan

Scan range: m/z 75-750 u Scan time: 60 microsec/u Collision energy: 13 eV Collision gas: Helium Mass resolution: Unit Precursor ion(s): m/z 277

Ions/transitions	Sample		Standard		Criteria	Dogult
	Area*	Ion Ratio	Area*	Ion Ratio	Tolerance <sup>#</sup>	Result
181	9489	0.449	128565	0.313	±25%	43.7%
257	1179	0.056	14673	0.036	±50%	56.5%
277	21115	1.000	411161	1.000	N/A	N/A

<sup>\*</sup>Peak area of the ion, % intensity compared to the most abundant ion

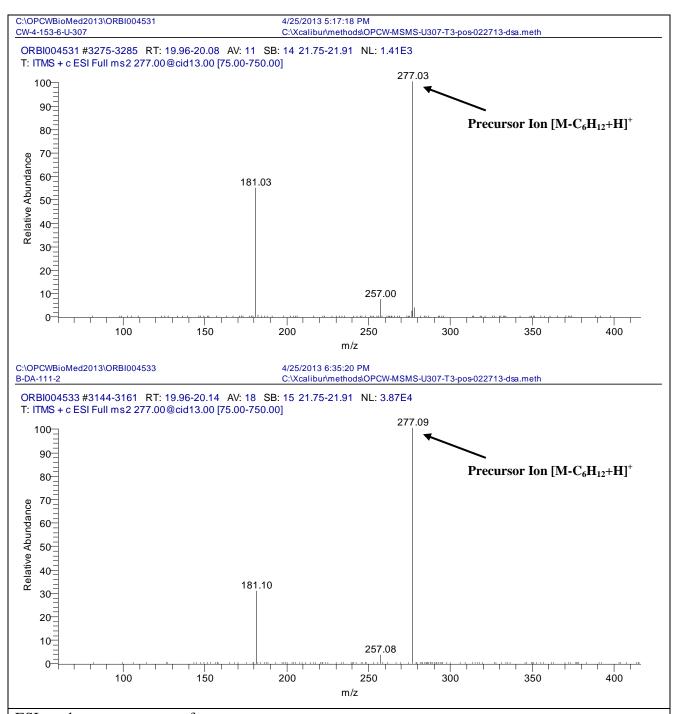
<sup>#</sup> Compare with the relative ion intensity of the standard: >50%:  $\pm 20\%$ ; >20% to 50%:  $\pm 25\%$ ; >10% to 20%:  $\pm 30\%$ ;  $\leq 10\%$ :  $\pm 50\%$ 



LC-MS/MS chromatograms supporting identification of **Chemical G-2 (Pentafluorobenzyl derivative)**; TIC.

Center: Chromatogram of the sample (aliquot code listed in header).

Bottom: Chromatogram of reference chemical of **Pentafluorobenzyl derivative of pinacolyl methylphosphonate.** 



Top: Chemical G-2 (Pentafluorobenzyl derivative) (aliquot code listed in header).

Bottom: Reference chemical of Pentafluorobenzyl derivative of pinacolyl methylphosphonate.